Lung cancer remains the leading cause of cancer mortality in the world with non-small cell lung cancer (NSCLC) accounting for 80% of cases (1). Conventional chemotherapeutic regimens only marginally improve the outcome of NSCLC patients at advanced stages of disease, with median survival time less than one year after diagnosis (2). Protein kinase activation by somatic mutation or chromosomal alteration is a common mechanism of tumorigenesis. The discovery of a number of these molecular alterations underlying lung cancer has led to uniquely targeted therapies with specific inhibitor drugs such as erlotinib and gefitinib for mutations in the epidermal growth factor receptor (EGFR), or crizotinib for the gene translocation resulting in the echinoderm microtubule associated protein like 4 (EML4)-anaplastic lymphoma kinase (ALK) oncogene (3-5). EML4-ALK was the first targetable fusion oncokine to be identified in 4-6% of lung adenocarcinomas. EML4-ALK generates a transforming tyrosine kinase with as many as nine different variants identified (6,7) and represents a novel molecular target in a small subset of NSCLCs. Patients with EML4-ALK positive tumors are characteristically younger age, female, and never to light smokers (5,8,9). The fusion gene has been observed predominantly in adenocarcinomas (4-7%) (5,9). Based on data from a phase I clinical trial which showed an overall response rate of 57% and a probability of progression-free...
survival at 6 months of 72%, crizotinib has been approved by the U.S. Food and Drug Administration for the treatment of NSCLC patients with ALK transforming rearrangements (5).

Similarly, proto-oncogene tyrosine-protein kinase ROS (ROS1) is an orphan receptor tyrosine kinase (RTK) that forms fusions and defines another clinically actionable oncogenic driver mutation in NSCLC (10). It has been recently reported that approximately 1.4% of NSCLCs harbor ROS1 rearrangements (11,12). Of the ROS1 fusion-positive tumors, 30% are known to harbor a recurrent translocation t[5;6][q32;q22], which creates the CD74 molecule, major histocompatibility complex, class II invariant chain (CD74)-ROS fusion kinase (13). In fact, ROS1 is evolutionarily related to ALK. Patients with ROS1 rearrangements are also significantly younger, more likely to be never-smokers and are more often diagnosed with the histological subtype of adenocarcinoma with wide distribution of tumor grade (11). Although ROS1 shares only 49% amino acid sequence homology with ALK in the kinase domains, several ALK inhibitors have demonstrated in vitro inhibitory activity against ROS1 (14). Recently, a report from investigators at the Massachusetts General Hospital Cancer Center has showed that ROS1-driven tumors can be treated with crizotinib and describes the remarkable response of one patient to crizotinib treatment (11). Interestingly, in this study ROS1 rearrangements were found to be mutually exclusive to ALK rearrangements (11). Preliminary results of a phase I trial of ROS1-positive advanced-stage NSCLC patients treated with crizotinib reported a response rate of 57% and a disease control rate of 79% at 8 weeks (15).

The discovery of new selective and potent inhibitors of ALK and ROS1 kinase raises the importance of using these drugs as a new method for treatment of ALK- and ROS1-derived lung cancer. This review focuses on the importance of targeting these proteins and describes the advances in optimizing more potent and selective ALK and ROS1 kinase inhibitors that have an optimal pharmacokinetic profile and the capacity to inhibit acquired resistant mutations. We aim to stimulate interest and encourage of researchers from different disciplines to learn about new therapeutic avenues following the development of compounds targeting ALK and ROS1 kinases with the aim of increasing survival to these lethal forms of lung cancer.

**Structural insights and computational simulations**

Receptor Tyrosine kinases (RTK) are transmembrane glycoproteins where the domain responsible to the tyrosine kinase activity is located in the cytoplasm. Although extracellular domain shows remarkable structural differences between TK families, the intracellular region is sensibly conserved.

Although a few years ago there was no resolved three-dimensional structure of ALK, similarity between its sequence permitted to predict its folding from a known RTK structure used as a template, using homology models. Thus, the human ALK receptor was modeled from mouse c-Abl (16), activated insulin receptor tyrosine kinase (InsR) (17,18) or insulin-like growth factor-1 receptor (IGF-1R) (19).

Fortunately, recently some crystal structures of the catalytic domain of ALK have been reported in literature at different resolution levels. All of them are available in the Protein Data Bank (PDB) (20) with ID entries 3L9P, 3LCS, 3LCT (Figure 1) (21).

Once the three-dimensional structure of the ALK receptor is available, biological processes related to its structure can be studied virtually, as for example substrate affinity (22), receptor autoactivation or resistant mutations. It can even be used as a structural template in order to predict the structure of RTK homologues by homology modeling (23).
However, crystallographic data should be taken carefully: the combination of crystallographic and biochemical studies reveals that the active conformation of ALK protein requires the phosphorylation of specific residues (24,25) and some ALK crystal structures published to date correspond to unphosphorylated proteins (26).

Additionally, some ALK molecular structures have also been resolved including a bound ligand (Table 1). These co-crystallized structures reveal the active site of the protein against one giving compound and describe the interaction within the protein-ligand complex. This information is relevant to assist structure-aided molecular design of ALK inhibitors (i.e. Structure-Based Drug Design, SBDD), providing valuable information about how to improve ALK selectivity.

From SBDD models rises the first approved drug for the treatment of ALK-positive NSCLC, crizotinib (29). By the time dianimopyrimidine (DAP) derivatives were consolidated as bioactive motives, great efforts were done in order to increase its selectivity (31), to optimize their activity (32) or to propose new scaffolds that mimic them. Evaluation of receptor-ligand interactions using docking techniques has become the most preferred SBDD method to study small-molecule ALK inhibitors, not only specific compounds as Novartis NVP-TAE684 (17) but also families including pyridine (33), pyrrolotriazine (19), 2-acyliminobenzimidazole (28), and tetrahydropyrido-2,3-b]pyrazine (34) derivatives. Besides specific chemical families, commercial or public chemical libraries can also be screened using docking techniques in order to identify new candidates (18,33).

Most of the SBDD results agree to consider the targeting of the ATP binding site of the tyrosine kinase domain as a good approach to design RTK inhibitors (35). The analysis of docking results may help to understand the inhibition mechanism, according to the interaction between candidates and the hydrophobic region and/or the gatekeeper. This helps to identify e.g., that preferred interactions of 2-aminopyridines within the ATP binding pocket include hydrogen bonds of pyridine and amino nitrogens with residues close to the gatekeeper (i.e. Met1184 and Glu1182) (34).

Ligand-based drug design (LBDD) has also been applied in the development of new ALK inhibitors. Structure-Activity Relationship (SAR) models correlate the chemical structure of inhibitors with their biological activity. They have been applied to screen series of piperidine carboxamides (27), 2-acyliminobenzimidazoles (28), macrocycles (36), 7-amino-1,3,4,5-tetrahydrobenzo[b]azepin-2-ones (37), 2,3,4,5-tetrahydro-benzo[d]azepines (38), 2,7-disubstituted pyrrolo[2,1-f][1,2,4]triazines (39), diaminocyclohexane methanesulfonamides (40) or tetracyclic derivatives (41,42), tetrahydropyrido[2,3-b]pyrazines (34), 3,5-diamino-1,2,4-triazole ureas (43), arloxy oxo pyrimidinones (44), identifying which substituents confer high ALK-response and proposing chemical modifications of hit compounds.

Far from what it seems, SBDD and LBDD are not exclusive. Most of published LBDD studies include modeling of candidates with the receptor using docking. LBDD and SBDD information can be combined to create pharmacophore models where chemical features (identified from LBDD methods) are used to generate structural keys that active ALK inhibitors must fulfill, taking into account receptor-ligand interactions (identified from SBDD methods) (19,45).

Since mechanisms to drug resistance have been experimentally related to mutations in the ALK amino acid sequence, many efforts have been done in order to obtain resolved crystal structures of ALK mutants (Table 2).

Ligand co-crystallizations are also available in mutated structures (mainly F1174L and R1275Q) (26), highlighting structural changes responsible of such behavior. Although resistance can be well correlated with structural changes in some cases (e.g., ALK\textsuperscript{F1174L}) (46), other mutations exhibit a more subtle behavior.

It is well know that ALK\textsuperscript{L1196M} mutants are crizotinib resistant (47). However, it is interesting to note that the comparison between co-crystal structure of crizotinib with ALK\textsuperscript{wt} (2XP2) (29) and ALK\textsuperscript{L1196M} (2YFX, McTigue M, 2012, unpublished data) reveals that L1196M mutation has little effect on crizotinib’s binding mode (Figure 2).

It has been recently demonstrated that at least four mutations could be involved in crizotinib’s resistance (Figure 3): L1196M acts as a gatekeeper and modifies crizotinib’s binding site by steric interference (49); G1202R and S1206Y are both located in the solvent-exposed region near to the binding site but G1202R blocks sterically the binding site whereas S1206Y could destabilize electrostatic interaction (50); 1151T insertion is furthest from N terminus Cα-helix but confers a high-level crizotinib resistance probably derived from the modification of ATP affinity (51). This knowledge has been used to design new compounds against gatekeeper mutants (52).

However, other mutations have also been identified. Most of them are abutting the Cα-helix and activation loop, becoming responsible for the modification of the kinase active site. On the basis of the crystal structures, these...
### Table 1 List of the available ligands co-crystallized with ALK receptor

<table>
<thead>
<tr>
<th>Molecular Structure</th>
<th>Ligand name</th>
<th>PDB ID (resolution)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td><img src="image1.png" alt="Image" /></td>
<td>piperidine-carboxamide</td>
<td>4DCE (2.03Å)</td>
<td>(27)</td>
</tr>
<tr>
<td><img src="image2.png" alt="Image" /></td>
<td>piperidine-carboxamide</td>
<td>4FNZ (2.60Å)</td>
<td>(26)</td>
</tr>
<tr>
<td><img src="image3.png" alt="Image" /></td>
<td>PHA-E429</td>
<td>2XBA (1.95Å)</td>
<td>(22)</td>
</tr>
<tr>
<td><img src="image4.png" alt="Image" /></td>
<td>NVP-TAE684</td>
<td>2XB7 (2.50Å)</td>
<td>(22)</td>
</tr>
<tr>
<td><img src="image5.png" alt="Image" /></td>
<td>acyliminobenzimidazole inhibitor 36</td>
<td>4FOD (2.00Å)</td>
<td>(28)</td>
</tr>
<tr>
<td><img src="image6.png" alt="Image" /></td>
<td>acyliminobenzimidazole inhibitor 1</td>
<td>4FOB (1.90Å)</td>
<td>(28)</td>
</tr>
<tr>
<td><img src="image7.png" alt="Image" /></td>
<td>acyliminobenzimidazole inhibitor 2</td>
<td>4FOC (1.70Å)</td>
<td>(28)</td>
</tr>
<tr>
<td><img src="image8.png" alt="Image" /></td>
<td>crizotinib</td>
<td>2XP2 (1.90Å)</td>
<td>(29)</td>
</tr>
<tr>
<td><img src="image9.png" alt="Image" /></td>
<td>CH5424802</td>
<td>3AOX (1.75Å)</td>
<td>(30)</td>
</tr>
</tbody>
</table>
Table 2 List of resolved structures of mutated ALK receptor

<table>
<thead>
<tr>
<th>Mutation</th>
<th>PDB ID</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>F1174L</td>
<td>2YJR (1.90Å)</td>
<td>McTigue M, 2012, unpublished data</td>
</tr>
<tr>
<td>F1174L catalytic domain</td>
<td>4FNW (1.75Å)</td>
<td>(26)</td>
</tr>
<tr>
<td>C1156Y</td>
<td>2YJS</td>
<td>McTigue M, 2012, unpublished data</td>
</tr>
<tr>
<td>L1196M</td>
<td>2YHV</td>
<td>McTigue M, 2012, unpublished data</td>
</tr>
<tr>
<td>L1196M + crizotinib</td>
<td>2YFX</td>
<td>McTigue M, 2012, unpublished data</td>
</tr>
<tr>
<td>R1275Q catalytic domain</td>
<td>4FNX (1.70Å)</td>
<td>(26)</td>
</tr>
<tr>
<td>R1275Q cd + benzoazole</td>
<td>4FNY (2.45Å)</td>
<td>(26)</td>
</tr>
</tbody>
</table>

Figure 2 Molecular structure of crizotinib co-crystallized with (A) ALK<sup>wt</sup> receptor, PDB ID: 2XP2; and (B) ALK<sup>L1196M</sup> mutant, PDB ID: 2YFX. Although Lys-Met exchange the binding mode of crizotinib remains unchanged. Structures analysis has been performed and rendered using MOE software (48).

mutations can be related with formation or alteration of hydrogen bonds (e.g., L1152R and C1156Y mutations) (53) or conformational changes (e.g., F1174L induces structural changes leading to an increment of ATP affinity which requires of irreversible inhibitors to block it) (35).

Thus, ligand co-crystallizations can not only help to the understanding of the structural basis of mutations related to drug resistance, but also to identify the binding mode of the second generation ALK inhibitors (30). When no crystallographic data is available, binding mode of new inhibitors with ALK mutants are computationally modeled using docking techniques (54) or molecular dynamic simulations (55). These interaction models provide a useful tool to screen drug candidates (after their validation with experimental evidences).

However, specificity for one receptor could be undesirable in a so complex biological context. For this reason, promiscuous inhibitors (e.g., dual inhibitors) have become gradually more interesting. When a set or library of knowing active ligands for one target is available, the specificity for the second target can be predicted by screening the previous library applying either SBDD or LBDD (56). In this respect, the ALK inhibitor TAE684 has been experimentally evaluated on ROS1, given the high homology between ALK and ROS1 human receptors (57–59).

Unfortunately, tridimensional structure of ROS1 receptor is not available and studies of ligand-receptor interactions have been always performed using ROS1 homology models using IGF-1R (60) or ALK itself (14). Homology models can be later used to study molecular interactions with ligands using docking methods in order to
elucidate their binding mode (60).

In contrast to homology models, other studies consider the structurally related MET receptor as a reference to find inhibitors which are able to interact with kinase domain. Given the receptor, homology docking studies on MET receptors could be therefore extended to structurally similar receptors like ROS1 (61). In fact, ALK inhibitors can also be used on ROS1 according to their similarity (62).

ALK and ROS1 small molecule inhibitors under development

In the last ten years a lot of efforts have been devoted to the development of compounds active against the ALK and ROS1 kinases. The principal reported inhibitors, the companies involved in the development, the core scaffold present in such compounds, the mutation against which the compounds are effective, their off-targets, and the clinical stage achieved are summarized in Table 3. On the other hand, the structures of the disclosed compounds are included in Figure 4.

ALK inhibition

One of the first discovered ALK inhibitors was NVP-TAE684, a small molecule with a dianilinopyridine scaffold which targets competitively the ATP in its binding site. Firstly, it was showed that NVP-TAE684 could block growth in cell lines and in a mouse model of ALCL (17). Besides to inhibit the proliferation of neuroblastoma and NSCLC cell lines, this ALK inhibitor reduced tumors expressing variants of ALK or EML4-ALK fusion proteins, confirming the oncogenic activity of the fusion kinase and consequently the therapeutic potential of targeted inhibitors (62,63). Despite several studies have reported the effectiveness of NVP-TAE684 against tumors induced by constitutively active ALK or against cell lines with ALK translocations and point mutations, this compound is not currently in any clinical trial (64-68).

Crizotinib (PF-02341066), which was originally developed to inhibit hepatocyte growth factor receptor (c-MET) but a few time later showed inhibitory activity against ALK, is an ATP-competitive small molecule like NVP-TAE684 (69,70). Crizotinib, with an aminopyridine as a core, was described for first time in 2007 and only three years later were reported the first promising clinical trials in NSCLC patients with EML4-ALK fusion genes (5). Phase I study concluded the benefits of the treatment with crizotinib of patients with advanced EML4-ALK-positive NSCLC. Among the 119 patients enrolled, after the treatment with crizotinib, 2 displayed a total recovery, 69 had a partial response and 31 were considered to have stable disease (71). The result of this inhibitory activity and the response of patients to the treatment with crizotinib, motivated its FDA approval under the trade name of Xalkori® (Pfizer). A Subsequent study in phase III trials reported crizotinib resistance in EML4-ALK positive NSCLC patients with some ALK mutations, especially secondary mutations, indicating a 64% overall survival in response to crizotinib treatment of EML4-ALK positive NSCLC patients after two years (49,53,72-74).

After the clinical trials of crizotinib, ALK was established as a drug target in cases of NSCLC, but the discovery of resistance related to mutations created the need to develop a second-generation of ALK inhibitors with the capability to overcome mutation-mediated drug resistance. Some pharmaceutical companies and research groups have reported different new promising candidate drugs to inhibit mutated ALK.

One of these second-generation ALK inhibitors is AP26113, whose chemical structure has not been disclosed but it is believed to be a member of a family compounds described in a patent of ARIAD based on a diaminopyrimidine structure bearing pendant phosphinoyl groups (75). This product is a multikinase inhibitor which shows more selectivity and inhibitory potency against ALK (IC50 =0.62 nM) than crizotinib (IC50 =3.6 nM). Apart from showing growth inhibition against EML4-ALK and nucleophosmin-ALK fusion gene (NPM-ALK) positive cells, the most interesting feature of AP26113 is that could inhibit some EML4-ALK mutated forms, therefore it could be a useful alternative in cases of crizotinib resistance. Currently, AP26113 is in phase I/II clinical trials (67,76-78).

In order to inhibit ALK mutations, another family of compounds was developed by Xcovery including X-276, X-376 and X-396 (79,80). There is not so much information in the literature about X-276 but it is considered a more selective and potent ALK inhibitor than crizotinib (81). X-376 and X-396 are members of the same family based on aminopyridazine scaffold, despite X-396 showed better results than X-376. X-396 was approximately 10-fold more potent than crizotinib in front of H3122 (EML4-ALK positive E13; A20 NSCLC), SU-DHL-1 (NPM-ALK positive E6a/b; A20 NSCLC), SU-DHL-1 (NPM-ALK positive), SY5Y (ALKF1174L) cell lines. As an example, in H3122 cell line, X-396 showed an IC50 value of 15 nM H3122.
Table 3 List of ALK inhibitors under development

<table>
<thead>
<tr>
<th>Chemical scaffold</th>
<th>Therapeutics</th>
<th>Company</th>
<th>ALK activity</th>
<th>ALK secondary mutations</th>
<th>Other targets</th>
<th>Clinical stage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Aminopyridine</td>
<td>Crizotinib PF-02341066 Xalkori®</td>
<td>Pfizer</td>
<td>24 nM</td>
<td>--</td>
<td>c-MET ROS1</td>
<td>FDA approved</td>
</tr>
<tr>
<td>Dianilinopyridine</td>
<td>NVP-TAE684</td>
<td>Novartis</td>
<td>&lt;10 nM</td>
<td>L1196M F1174L</td>
<td>IR, IGF-1R, FLT3, TIE2 LRRK2, ROS1</td>
<td>Not a clinical candidate</td>
</tr>
<tr>
<td>Structure undisclosed</td>
<td>AP26113</td>
<td>ARIAD</td>
<td>0.62 nM</td>
<td>L1196M F1174C I1171T F1245C E1210K S1206R G1269S</td>
<td>Multiple kinases</td>
<td>Phase I/II</td>
</tr>
<tr>
<td>Structure undisclosed</td>
<td>X-276</td>
<td>Xcovery</td>
<td>--</td>
<td>--</td>
<td>--</td>
<td>preclinical</td>
</tr>
<tr>
<td>aminopyridazine</td>
<td>X-376</td>
<td>Xcovery</td>
<td>--</td>
<td>--</td>
<td>--</td>
<td>preclinical</td>
</tr>
<tr>
<td>Structure undisclosed</td>
<td>Aminopyridazine</td>
<td>X-396</td>
<td>Xcovery</td>
<td>&lt;0.4 nM</td>
<td>L1196M C1156Y</td>
<td>--</td>
</tr>
<tr>
<td>Tetracyclic indole</td>
<td>CH5424802 AF-802</td>
<td>Chugai</td>
<td>1.9 nM</td>
<td>L1196M F1174L R1275Q C1156Y</td>
<td>GAK, LTK</td>
<td>Phase I/II</td>
</tr>
<tr>
<td>Pyrrolopyrimidin</td>
<td>GSK1838705A</td>
<td>GlaxoSmithKline</td>
<td>0.5 nM</td>
<td>--</td>
<td>IR</td>
<td>Preclinical</td>
</tr>
<tr>
<td>Triazinediamine</td>
<td>ASP3026</td>
<td>Astellas</td>
<td>--</td>
<td>--</td>
<td>ROS1</td>
<td>Phase I</td>
</tr>
<tr>
<td>Indolocarbazole</td>
<td>CEP-14083</td>
<td>Cephalon</td>
<td>2 nM</td>
<td>--</td>
<td>IR, VEGFR2, TIE2, DLK</td>
<td>Not a clinical candidate</td>
</tr>
<tr>
<td>Indolocarbazole</td>
<td>CEP-14513</td>
<td>Cephalon</td>
<td>4 nM</td>
<td>--</td>
<td>IR, VEGFR2, TIE2, DLK</td>
<td>Not a clinical candidate</td>
</tr>
<tr>
<td>Tetrahydropyrazine</td>
<td>--</td>
<td>Cephalon</td>
<td>10 nM</td>
<td>--</td>
<td>--</td>
<td>preclinical</td>
</tr>
<tr>
<td>Diaminopyridine</td>
<td>CEP-28122</td>
<td>Cephalon</td>
<td>1.9 nM</td>
<td>F1174L R1275Q</td>
<td>--</td>
<td>Phase I</td>
</tr>
<tr>
<td>Structure undisclosed</td>
<td>CEP-37440</td>
<td>Cephalon/Teva</td>
<td>--</td>
<td>--</td>
<td>--</td>
<td>Phase I</td>
</tr>
<tr>
<td>Pyrrolotriazine</td>
<td>Compound 32</td>
<td>Cephalon</td>
<td>6 nM</td>
<td>--</td>
<td>--</td>
<td>preclinical</td>
</tr>
<tr>
<td>Tetraazatetracyclo-docosanonaene</td>
<td>Macrocyclic 2m</td>
<td>Cephalon</td>
<td>0.5 nM</td>
<td>--</td>
<td>--</td>
<td>preclinical</td>
</tr>
<tr>
<td>Pyrrolopyrazole</td>
<td>PHA-E429</td>
<td>Nerviano Medical</td>
<td>91 nM</td>
<td>--</td>
<td>Multiple kinases</td>
<td>preclinical</td>
</tr>
<tr>
<td>Indazole</td>
<td>NMS-E628</td>
<td>Nerviano Medical</td>
<td>55 nM</td>
<td>L1196M C1156Y</td>
<td>IGF-1R, Aurora B</td>
<td>preclinical</td>
</tr>
<tr>
<td>Structure undisclosed pyridone</td>
<td>CRL151104A</td>
<td>Chembridge St Jude</td>
<td>9.75 nM</td>
<td>F1174L R1275Q</td>
<td>--</td>
<td>preclinical</td>
</tr>
<tr>
<td>Pyridone</td>
<td>Pyridone 1</td>
<td>Chembridge St Jude</td>
<td>380 nM</td>
<td>--</td>
<td>--</td>
<td>preclinical</td>
</tr>
<tr>
<td>Structure undisclosed</td>
<td>WZ-5-126</td>
<td>--</td>
<td>3.4 nM</td>
<td>--</td>
<td>--</td>
<td>preclinical</td>
</tr>
<tr>
<td>Structure undisclosed</td>
<td>LDK378</td>
<td>Novartis</td>
<td>0.15 nM</td>
<td>--</td>
<td>ROS1</td>
<td>Phase I</td>
</tr>
<tr>
<td>2,4-pyrimidinediamine</td>
<td>3-39</td>
<td>Novartis</td>
<td>--</td>
<td>--</td>
<td>--</td>
<td>preclinical</td>
</tr>
<tr>
<td>Pyridoisoquinoline</td>
<td>F91873 and F91874</td>
<td>Institut de Recherche Pierre Fabre</td>
<td>--</td>
<td>--</td>
<td>Multiple kinases</td>
<td>preclinical</td>
</tr>
<tr>
<td>Structure undisclosed</td>
<td>TSR-011</td>
<td>Tesaro</td>
<td>--</td>
<td>--</td>
<td>--</td>
<td>preclinical</td>
</tr>
</tbody>
</table>

c-MET, hepatocyte growth factor receptor; IR, insulin receptor; IGF-1R, insulin-like growth factor 1 receptor; FLT3, FMS-like tyrosine kinase 3; TIE2, angiopoietin-1 receptor; LRRK2, leucine-rich repeat kinase 2; GAK, cyclin G-associated kinase; LTK, leukocyte tyrosine kinase; VEGFR2, vascular endothelial growth factor receptor 2; DLK, dual leucine zipper kinase
Figure 4 Structures of reported ALK and ROS1 inhibitors
(EML4-ALK positive NSCLC) cell lines against 180 nM of crizotinib. Furthermore X-396 displayed good inhibition in crizotinib resistant cell lines (Ba/F3-EML4-ALK1196M, IC_{10} =106 nM; Ba/F3-EML4-ALK^F1174L, IC_{10} =48 nM). Thus, X-396 initiated phase I clinical trials in June 2012 (82).

CH5424802, also known as AF-802, is a tetracyclic indole which has a high ALK<sup>wt</sup> inhibitory activity in <i>in vitro</i> assays (IC_{10} =1.9 nM) as well in front of mutated ALKs, such as ALK^L1196M (K<sub>i</sub> =1.56 nM), ALK^F1174L (IC_{10} =1.0 nM) and ALK^R1275Q (IC_{10} =3.5 nM). These results were also reproduced in <i>vivo</i> with the treatment in different cell lines: H2228 (EML4-ALK positive E6a/b;A20; IC_{10} =53 nM), KARPAS-299 (NPM-ALK positive ALCL; IC_{10} =3.0 nM), SR-786 (IC_{10}=6.9 nM), NB-1 (ALK amp, IC_{10} =4.5 nM); KELLY (IC_{10} =62 nM) and Ba/F3 (EML4-ALK<sup>L1196M</sup>) that allowed that CH5424802 is a potent inhibitor for a therapy with capacity to overcome the acquired resistance to crizotinib (30,41,83). Due to these promising results, CH5424802 is in phase II/II of clinical trials.

GSK1838705A, developed by GlaxoSmithKline and currently in preclinical phase, contains a pyrrolopyrimidine scaffold and has showed to be selective IGF-1R, insulin receptor (IR) and ALK inhibitor (IC<sub>90</sub> =1.2, 2 and 0.5 nM respectively). Furthermore, the inhibition of the proliferation of different ALCL cell lines, such as L-82 (IC_{10} =24 nM), SUP-M2 (IC_{10} =28 nM), SU-DHL-1 (IC_{10} =31 nM), Karpas-299 (IC_{10} =52 nM) and SR-786 (IC_{10} =88 nM), has also been described. Besides the potent inhibition of ALK by GSK1838705A, such compound also inhibit cell lines harboring ALK fusion genes in different ALCL cell lines expressing NPM-ALK (EC_{50} =24-88 nM) and in H2228 NSCLC cells expressing EML4-ALK (IC_{10} =191 nM). In addition, it was proved that GSK1838705A inhibits the EML4-ALK phosphorylation (84,85).

There are not so many details about ASP3026, a triazinediamine developed by Astellas which is in Phase I in clinical trials. This compound showed potent and selective activity against EML4-ALK driven tumors with gatekeeper mutation, therefore it is able to overcome crizotinib resistance (86).

Based on the structure of two natural products, staurosporine and 7-hydroxystaurosporine which are able to inhibit ALK (IC_{10} =150 nM and 5 μM respectively in the presence of 30 μM ATP in an ELISA-based ALK assay) (87), Cephalon developed some compounds targeted to inhibit ALK. CEP-14083 and CEP-14513 have showed ATP-competitive activity in ALK, displaying IC_{50} values in enzymatic assays of 2 and 4 nM, respectively, and 10-30 nM in cellular assays (88). The capability of CEP-14083 to inhibit cell lines and animal models harboring ALK alterations was also described (89). Furthermore, CEP-14083 is able to inhibit other kinases, such as IR, vascular endothelial growth factor receptor 2 (VEGFR2), angiopoietin-1 receptor (TIE2), and dual leucine zipper kinase (DLK) but, due to unfavorable physicochemical properties, CEP-14083 and CEP-14513 were discarded for <i>vivo</i> studies (88). Then, Cephalon developed a second generation of ALK inhibitors with different scaffolds, some of them being currently in preclinical studies. Thus, they reported tetrahydroprazines with IC_{10} around 10 nM and 150 nM in enzyme and in Karpas-299 cell line, respectively (34). They also described 2,4-diaminopyrimidines, the most representative compound is CEP-28122, showing a high selectivity (600-fold with respect to IR, a closely related kinase family member) and a high potency (IC_{10} =1.9 and 20 nM in enzyme and in Karpas-299 cell line, respectively). CEP-28122 also induced growth inhibition in NPM-ALK positive ALCL and EML4-ALK positive NSCLC tumor xenografts in mice. In addition, inhibited growth of neuroblastoma cell lines harboring ALK activating mutants, such as F1174L in NB-1643 cells and R1275Q in SH-SY5Y cells, but not in cell lines which expresses ALK<sup>wt</sup> (SKNAS cells) (32,37,90). Cephalon also described pyrrolotriazines among which compound 32 displayed an IC_{10} =6 nM in an enzymatic assay and 100 nM against NPM-ALK cells, showing high selectivity in a test against 256 kinases (39). Finally, they reported tetrazatatetracyclo docosanaena macrocycles, Macrocycl 2m showing a high inhibitory potency <i>in vitro</i> (IC_{10} =0.5 nM) as well as in a cell-based assay (IC_{50} =10 nM) and high selectivity (173-fold with respect to IR) (36).

PHA-E429 is a compound developed by Nerviano with a pyrrolopyrazole scaffold which is considered as a multikinase inhibitor that also is able to inhibit ALK (IC_{90} =91 nM) (51,91). NMS-E628, an indazole compound, developed by the same company, inhibits ALK (IC_{50} =55 nM against NMP-ALK cells) but also IGF-1R and Aurora B (92). NMS-E628 has showed a high efficiency inhibiting the growth of ALCL cells and NSCLC cells bearing EML4-ALK (H2228) rearrangement and in addition was able to overcome the L1196M and C1156Y mediated TKI resistance (93,94).

CRL151104A, developed by ChemBridge Research Laboratories and St Jude Children’s Research Hospital, whose structure is not available, is a third-generation ATP
competitor pyridine compound with capacity to block the cellular phosphorylation. This compound showed a high activity against ALK in vitro (IC<sub>50</sub> = 9.75 nM) and also in an in vivo assay (IC<sub>50</sub> ≤ 100 nM and 2.5 μM against NPM-ALK positive and NPM-ALK negative cells, respectively). CRL151104A has also demonstrated the capability to overcome F1174L and R1275Q ALK mutations in neuroblastoma cell lines (95). There is not so much information about a pyridone compound developed by ChemBridge Research Laboratories and St Jude Children’s Research Hospital which is able to inhibit ALK in an enzymatic assay (IC<sub>50</sub> = 380 nM) and in cell-based assays (IC<sub>50</sub> = 18.0 μM and 750 nM against BaF3/NPM-ALK and Karpas-299/NPM-ALK ALCL cell lines, respectively) (33). WZ-5-126 is a potent ALK small molecule inhibitor with an IC<sub>50</sub> of 3.4 nM in vitro, capable of inhibiting the growth of two ALK positive NSCLC cell lines (62).

Novartis has also been involved in the research of ALK inhibitors such as LDK-378, a potent and selective candidate (IC<sub>50</sub> = 0.15 nM). This compound, in Phase I trials nowadays, is able to inhibit the growth of genetically abnormal ALK-driven NSCLC tumors (96). Furthermore, compound 3-39 described in a Novartis patent, including a 2,4-pyrimidinediamine scaffold, showed brilliant results against EML4-ALK transgenic mouse models (63, 97).

On the other hand, F91873 and F91874 are pyridoisoquinolines with multikinase inhibitory activity described by Institut de Recherche Pierre Fabre which are able to inhibit ALK and probably behave as non-ATP competitors (98).

Another ALK inhibitor in preclinical development is TSR-011, developed by Tesaro, but unfortunately the information about it is very scarce (99).

Finally, some other compounds have been reported that are not able to directly inhibit ALK but are able to inhibit the tumor growth in ALK-driven NSCLC models, such as EML4-ALK<sub>L1196M</sub>, therefore being capable to overcome crizotinib resistance. Two examples of such compounds are the Hsp90 inhibitors retaspimycin (IPI-504) and ganetespib (STA-9090) (67, 100-102).

Some kinase inhibitors were assayed against ROS1 and one of them, staurosporine, showed high inhibitory activity (IC<sub>50</sub> = 0.9 nM) (61). On the other hand, several heterocyclic compounds, such as AST-487, PP 2, AG 1487, PDGFR-I-III and D-64406, showed moderate to low inhibitory activities (IC<sub>50</sub> = 1,700, 5,200, 13,600, 48,000 and 365,000 nM respectively) (103-105).

Due to the high homology between the kinase domains of ROS1 and ALK, some ALK inhibitors were assayed against ROS1-driven cells and tumors. NVP-TAE684 showed in vitro activity against HCC78 cell lines expressing ROS1 and inhibition of signaling downstream of ROS1 inducing apoptosis in BaF3/FIG-ROS positive cells (IC<sub>50</sub> = 10 nM) (57, 79). Furthermore, a computational study of NVP-TAE684 showed its higher affinity for the ROS1 kinase domain with respect to the ALK kinase domain (14).

Apart from being the first ALK inhibitor approved by FDA, crizotinib is also able to inhibit ROS1 with an IC<sub>50</sub> value of 1.7 nM in an in vitro assay. This high inhibition in an enzymatic-based assay was not confirmed in a cell-based assay (IC<sub>50</sub> = 1.4 μM against the HCC78 ROS1-rearranged NSCLC cell line) (79). The inhibition of the ROS1 phosphorylation by crizotinib in the HCC78 cell line in a moderate manner was also described (106). Although only 2% of NSCLC cases harbor ROS1 rearrangements, it is described the case of a patient having a ROS1 positive tumor (without EGFR mutations nor ALK rearrangements) who did not respond to erlotinib treatment (an EGFR inhibitor) but was totally recovered in 12 weeks after crizotinib treatment. This case is the proof that ROS1 is a prime target for the development of new inhibitors for the treatment of NSCLC (11).

Some other ALK inhibitors were assayed against ROS1 with promising results. Thus AP26113 (IC<sub>50</sub> = 1.9 nM) (67) and WZ-5-126 (IC<sub>50</sub> = 8.2 nM) (51), together with crizotinib and ASP3026, have entered clinical trials (59).

Finally, there are only two pyrazole ROS1 selective compounds (KIST301072 and KIST301080), which showed good ROS1 inhibitory activity (IC<sub>50</sub> = 199 and 209 nM, respectively) when tested against 45 kinases (60, 106, 107).

**Conclusions**

Traditionally, depending on the type of tumor, therapeutic approaches include different combinations of surgery, radiation therapy, and chemotherapy. However, alternative therapies using receptor tyrosine kinase (RTK) as targets started to be introduced in the beginning of this century. In
the case of lung cancer, such approach has led to discovery of a number of these targeted therapies with specific inhibitor drugs such as erlotinib and gefitinib for mutations in the epidermal growth factor receptor (EGFR). However, in a 4-7% of the of lung adenocarcinomas these treatments are ineffective due to the presence of ALK and/or ROS1 rearrangements. This problem has been partially overcome due to the development of crizotinib (Xalkori®, Pfizer) but the discovery of some crizotinib resistant ALK mutations has forced the research of new inhibitors. Furthermore, the implication of ROS1 protooncogene in this kind of tumors has rendered the situation even more complicated. The present review has tried to show the importance of ALK and ROS1 as combined targets for the development of new treatments for non-small cell lung cancer (NSCLC) when the most common approaches fail and the efforts carried out in this field to date.

Acknowledgements

Disclosure: The authors declare no conflict of interest.

References


